# 12-HYDROXYTOVAROL AND DERIVATIVES FROM THAPSIA VILLOSA VAR. MINOR

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Abstract—The new germacrane esters 12-angeloyloxy- and 12-hydroxy-8-O-angeloyltovarol and 12-angeloyloxy-8-O-angeloylshiromodiol were isolated from T. villosa var. minor, and their structures established by spectral data and chemical correlations. The terpenoids geranyl acetate,  $\delta$ -cadinene,  $\gamma$ -cadinene,  $\gamma$ -muurolene,  $\beta$ -caryophyllene and caryophyllene oxide were also isolated from the aerial parts.

## INTRODUCTION

In the course of our study on the chemical components from Thapsia species [1-4] we have described the isolation of some germacrane esters related to tovarol [=  $(6S,7R,8S,4E,9E)\cdot1(10),4$ -germacradiene-6,8-diol] and shiromodiol  $(4\beta,5\beta$ -epoxy derivative). We would like to report here the isolation of two new 12-hydroxytovarol angelates 1 and 2 from the roots of T. villosa var. minor and a new 12-angeloyloxyshiromodiol derivative 14 as well as other known terpenoids from the umbellas of this plant.

## RESULTS AND DISCUSSION

The benzene extract from the roots of the plant afforded after chromatography pure 1 and 2. Both substances showed IR spectra nearly identical and very similar to that of 8-O-angeloyltovarol previously isolated from the same plant [2]. The IR spectra were indicative of the presence of hydroxyl groups (3400 cm<sup>-1</sup>), conjugated ester (1700, 1230 cm<sup>-1</sup>) and double bonds (1640, 850 cm<sup>-1</sup>). The <sup>1</sup>H NMR spectra of 1 and 2 showed the presence of two olefinic protons, two vinyl methyl groups as well as a secondary methyl group. Compound 1 showed also signals of two angeloxy groups but 2 displayed signals of only one angeloxy group. The alkaline hydrolysis of both 1 and 2 gave the same triol 3 which on standing at room temperature overnight with acetic anhydride-pyridine gave the triacetate 4. The <sup>1</sup>H NMR spectra of 3 and 4 were quite similar to those of tovarol 5 and its diacetate 6, respectively. However, tovarol showed signals of an isopropyl group while 1 to 4 showed only one 3H doublet and signals of a CH2OR group. These data suggested that the third oxygenated group should be placed on C-12. Moreover, the coupling constants displayed by the acetates 4 and 6 are analogous and both showed high specific rotations with the same sign (4:  $[\alpha]_D$  $-71.0^{\circ}$ ; 6:  $[\alpha]_{D} - 101.5^{\circ}$ ). This suggested that the configuration of both compounds was the same except for the new C-11 chiral centre.

To establish the configuration at C-11 the isopro-

7 R: OH 8 R: OAc

9 R: =0

10

pylidene derivatives of triol 3 were synthesized. As the hydroxyl groups attached to C-6 and C-8 are trans the 6,12 and 8,12 dioxepanes were the only expected isopropylidene derivatives. In fact only two products were obtained, 7 and 10. Product 7 was acetylated yielding 8. This showed the H-8 multiplet deshielded and located the free hydroxyl group at C-8. Assuming a conformation for the cyclodecane ring 13D5, 1D14 [5] analogous to that shown by shiromodiol [6], as suggested by the observed coupling constants, the oxepane ring may adopt the halfchair conformation I or II as the half-boat conformations are highly puckered because of the steric interactions of the isopropylidene group with the cyclodecane substituents. Although both conformations allow the same conclusions with respect to the C-11 configuration, I must be the more stable conformation for 7 as deduced from the deshielding of one of the C-12 protons. This deshielding could be caused by the n electrons of the C-6 oxygen being spatially close to the H-12 in conformation I. Also, the small coupling constants between the C-12 protons with H-11 (H-12 $\beta$ :  $\delta$ 4.00, brd, J = 12 Hz; H-12 $\alpha$ :  $\delta$ 3.25, br

н	1	2	3	4	7	8	9	10	13	14
1	5.10 m	5.10 m	5.00 m	5.10 m	5.00 m	5.20 m	4.90 m	4.85 m	5.15 m	5.28 m
2, 3	2.12 br s	2.15 br s	2.15 br s	2.20 br s	2.12 br s	2.18 br s	2.10 br s	2.10 br s	2.15 br s	
5	5.10 m	5.10 m	5.00 m	5.10 m	5.00 m	5.20 m	4.90 m	4.85 m	4.90 br d (9)	2.82 br d (7)
6	5.10 m	5.10 m	4.70 br d	5.40 br d	4.60 br d	4.60 br d	4,55 br d	4.85 m	4.40 dd	3.42 br d
			(7.5)	(7)	(7)	(7)	(7.5)		(9, 6)	(7)
8	4.20 m	4.50 br t (7)	4.10 m	5.10 m	4.15 m	5.20 m	,	4.00 m	5.15 m	5.28 m
12	4.20 m	3.50 m	3.55 m	4.15 dd	4.15 br d	3.92 br d	4.25 br d	4.00 m	3.80 dd	4.15 ABm
				(10, 3)	(12)	(12)	(12)		(9, 7)	
				3.85 dd	3.25 br d	3.25 br d	3.10 br d	3.25 ₺	3.05 dd	
				(10, 4)	(12)	(12)	(12)	(12)	(9, 7)	
13	1.15 d	1.10 d	0.97 d	1.15 d	1.10 d	1.15 d	1.07 d	1.00 d	1.10 d	1.20 d
	(6.5)	(6.5)	(6.5)	(6.5)	(6.5)	(6.5)	(6.5)	(6.5)	(6.5)	(6.5)
14	1.62 s	1.68 s	1.60 s	1.70 s	1.60 s	1.65 s	1.50 s	1.60 s	1.67 s	1.80 s
15	1.38 s	1.48 s	1.45 s	1.50 s	1.45 s	1.48 s	1.50 s	1.40 s	1.46 s	1.25 s
Ang	6.05 br q	6.00 br q							5.95 br q	6.05 br q
	5.95 br q (6)	(6)							(6)	5.95 br q (6)
	1.96 br d	1.96 br d							1.96 br d	1.95 m
	(6)	(6)							(6)	
	1.90 br s	1.90 br s							1.90 br s	1.90 br s
Ac				2.01 s		1.97 s				
				2.00 s						
				1.95 s						
gem-Mc <sub>2</sub>					1.30 s	1.22 s	1.30 s 1.19 s	1. <b>40</b> s		

Table 1. 1H NMR spectral data for 12-hydroxytoyarol derivatives\*

<sup>\*</sup>Recorded at 60 MHz in CDCl<sub>3</sub> (3, 4, 10, 14) or in CCl<sub>4</sub>.  $\delta$  scale in ppm relative to TMS and J in Hz (in parentheses).

d, J=12 Hz) are in agreement with the conformation shown in I. The Eu(fod)<sub>3</sub> induced shift on the NMR signal for Me-13 of compound 7 is small as compared with the H-8 induced shift ( $\Delta\delta_{\text{Me-13}}$  0.3 vs.  $\Delta\delta_{\text{H-8}}$  2.8) and this is against the R-configuration for C-11 in which the Me-13 group points in the same direction as the hydroxyl group in both conformations I and II. In consequence we propose the S-configuration for C-11.

This assignment is also in agreement with the observed chemical shift for Me-13 in the oxidation product 9. Assuming for this compound a ring conformation like that shown in II, the Me-13 is placed below the carbonyl plane if the C-11 configuration is R, whereas if the configuration is S this methyl group is placed away from the carbonyl group. As the doublet of Me-13 is not significantly shielded (7:  $\delta$ 1.10; 9:  $\delta$ 1.07) it is also concluded that the configuration for C-11 must be S.

A germacradiene lactol named 'hallerin' (11) has been isolated recently from Laserpitium halleri [7]. The absolute configuration of the chiral carbon atoms C-6, C-7 and C-8 of 11 is the same that the previously proposed for our tovarol derivatives. To confirm the structure of our substances, an authentic sample of the reduction product from hallerin-lactone 12 was requested. We found that the <sup>1</sup>H NMR and IR spectra, as well as the mp and/or  $[\alpha]_D$  for compounds 3 and 4 were identical to those of the triol and triacetate synthesized from natural hallerin. Consequently the configuration shown in 3 for 12-hydroxytovarol was confirmed

The site where the angeloxy groups are attached in the natural tovarol esters 1 and 2 was deduced from their <sup>1</sup>H NMR spectra as compared with those of the derivatives 3–8. The hydrolysis of diangelate 1, caused an upfield shift for the signals of the H-12 protons ( $\Delta\delta$ 0.6). The doublet at  $\delta$ 5.40 in triacetate 4 assigned to H-6 is deshielded as expected with respect to the same signal in triol 3 ( $\Delta\delta$ 0.7 ppm) but the H-6 signal in 1, partially overlapped near 5 ppm, is shielded after hydrolysis by 0.3 ppm only. This suggested that the non-esterified hydroxyl group is that placed on C-6 as shown in formula

Compound 2, esterified by one angeloxy group, showed the same chemical shifts as compound 1 for H-6 and H-8, but the H-12 protons were shielded with respect to 1

14

**11** R ОН **12** R =0

13 R H

 $(\Delta\delta0.7 \text{ ppm})$ . These data are indicative that the angeloxy group in this case must be attached to C-8. This substitution pattern was also evidenced by cyclization. Treatment of 2 with TsCl gave the tetrahydrofurane 13.

The benzene extract from the umbellas of the plant was also studied. The main component was identified as geranyl acetate. The known sesquiterpenoids  $\delta$ - and  $\gamma$ -cadinene,  $\gamma$ -muurolene,  $\beta$ -cariophillene and  $\beta$ -cariophillene oxide were also identified, as well as falcarindiol, sitosterol,  $\beta$ -amyrin and glycerides (GC methyl esters: linoleate 60% oleate 13%, linolenate 7%.

From the same extract a new germacrane ester 14 was isolated. The IR spectrum of 14 was similar to that of 8-O-angeloylshiromodiol [2] but the  $^{1}H$  NMR spectrum showed in this case the presence of two angelate groups, one epoxydic proton (2.82, 1H, d, J=6 Hz), one methyl doublet (1.20, 3H, d, J=6 Hz) and signals for CHOAng (5.28, 1H, m) and CH-CH<sub>2</sub>OAng (4.15, 2H, m). All these data led us to assign the structure shown in 14 for this new toyarol derivative.

To confirm the structure proposed for 14, the natural tovarol (1) was epoxidized with one mol of m-CPBA to give a monoepoxide identical in all respects to compound 14

## EXPERIMENTAL

Isolation. Plant material was collected, extracted and fractionated as previously reported [2]. Silica gel chromatography (I kg) of the roots neutral fraction (236 g) with hexane-Et<sub>2</sub>O mixtures of increasing polarity was carried out. Fractions containing 1 (12 g) were further chromatographed on silica gel with hexane-EtOAc (9:1) and on silica gel H-60 (60 g,  $C_6H_6$ -EtOAc, 97:3) yielding pure 1 (760 mg). Fractions containing 2 (19 g) yielded after chromatography on silica gel eluted with hexane-EtOAc (7:3),  $C_6H_6$ -EtOAc (8:2) and hexane-Et<sub>2</sub>O (6:4), pure 2 (490 mg).

The defatted umbellas extract (31.9 g) was chromatographed (silica gel, 500 g) with hexane— $Et_2O$  mixtures of increasing polarity. The less polar fractions (1.2 g) were further chromatographed on  $10^{\circ}$ , AgNO<sub>3</sub>—silica gel yielding  $\delta$ -cadinene (40 mg),  $\gamma$ -muurolene (60 mg),  $\gamma$ -cadinene (20 mg) and  $\beta$ -caryophyllene (200 mg). The next fractions contained geranyl acetate (10.3 g), caryophyllene oxide (98 mg) and an oily product (290 mg) which after chromatography on silica gel H-60 (30 g) with hexanc-  $Et_2O$  (8:2) yielded pure 14 (166 mg).

12-Angeloyloxy-8-O-angeloyltovarol (1). Oily,  $[a]_D = 132.0^\circ$  (CHCl<sub>3</sub>; c 7.3). IR  $v_{max}^{lim}$  cm<sup>-1</sup>: 3400, 1700, 1640, 1380, 1220, 1150, 1070, 1030, 1000, 850. EIMS (probe) 70 eV, m/z (rel. int.): 218 [M - 2AngOH]\* (10), 203 [M - 2AngOH - Me]\* (3), 200 [M - 2AngOH - H<sub>2</sub>O]\* (10), 203 [M - AngOH - Me]\* (3), 200 [M - 2AngOH - H<sub>2</sub>O]\* (4), 169 [M - AngOH - iso - PrOAng - H<sub>2</sub>O]\* (15), 134 (25), 121 (10), 119 (10), 107 (40), 105 (20), 93 (60), 83 (100), 57 (50), 55 (50), 43 (20), 41 (20).

12-Hydroxy-8-O-angeloyltovarol (2). Oily. IR v <sup>5lm</sup>c cm<sup>-1</sup>: 3400, 1710, 1650, 1450, 1390, 1240, 1160, 1080, 1050, 1000, 850.

12-Hydroxytovarol (3). Compounds 1 or 2 hydrolysed overnight with 2 N NaOH MeOH yielded crystalline 3. Mp 164–165° (Et<sub>2</sub>O). [ $\alpha$ ]<sub>D</sub> =88.2° (EtOH; c 1.9) 1R  $\nu$ <sup>nuyol</sup> cm<sup>-1</sup>: 3300, 2950, 1670, 1380, 1260, 1060, 1020, 980, 860, 840, 760, 680. EIMS (probe) 70 eV, m/z (rel. int.): 236 [M]\* (4), 221 [M =15]\* (16), 218 [M = 18]\* (10), 177 (15), 159 (40), 121 (40), 107 (60), 93 (95), 81 (100), 69 (70), 55 (50), 41 (40).

Triacetate 4. Acetylation of 3 (170 mg) in Ac<sub>2</sub>O-pyridine yielded 4 (190 mg), which was purified by CC on silica gel with hexane-Et<sub>2</sub>O (8:2) (110 mg). Oily,  $[\alpha]_D = 78.0^{\circ}$  (CHCl<sub>3</sub>; c 4.3).

IR  $v_{\text{max}}^{\text{lim}}$  cm<sup>-1</sup>: 2950, 1740, 1660, 1450, 1380, 1250, 1130, 840. EIMS (probe) 70 eV, m/z (rel. int.): 260 [M - 2AcOH]\* (4), 218 [M - C<sub>2</sub>H<sub>2</sub>O]\* (2), 200 [M - 3AcOH]\* (10), 185 (85), 159 [M - 2AcOH - iso-PrOAc]\* (100), 93 (85), 43 (90).

6,12-Isopropylidene derivative 7. Reaction of 3 (95 mg) in 4 ml Me<sub>2</sub>CO with 2,2-dimethoxypropane (1 ml) and a trace of p-TsOH afforded after work up a crude oil which was chromatographed on silica gel (10g; hexane-Et<sub>2</sub>O, 8:2), to yield 7 (63 mg) and 10 (21 mg). Compound 7: Oily. [ $\alpha$ ]<sub>D</sub> = 118.1° (CHCl<sub>3</sub>; c 1.6). IR  $\nu_{\max}^{\text{tim}}$  cm<sup>-1</sup>: 3000, 2950, 1670, 1460, 1390, 1230, 1170, 1060, 1000, 950, 900, 870, 850, 800, 710. EIMS (probe) 70 eV, m/z (rel. int.): 264 [M - 2Me]\* (4), 236 [M - C<sub>3</sub>H<sub>6</sub>O]\* (10), 227 [M - C<sub>3</sub>H<sub>6</sub>O - Me]\* (1), 218 [M - C<sub>3</sub>H<sub>6</sub>O - H<sub>2</sub>O]\* (1), 206 (10), 187 (10), 159 (25), 152 (25), 43 (100).

Acetate 8. Acetylation of 7 (70 mg) under the usual conditions (Ac<sub>2</sub>O-pyridine, room temp.) yielded 8 (72 mg). Oily.  $[\alpha]_D = 104.6^\circ$  (CHCl<sub>3</sub>; c 3.6). IR  $v_{max}^{time}$  cm<sup>-1</sup>: 1740, 1660, 1370, 1240, 1210, 1050, 1020, 1000, 860, 840. EIMS (probe) 70 eV m/z (rel. int.); 278  $[M-C_3H_6O]^+$  (20), 276  $[M-AcOH]^+$  (20), 218  $[M-AcOH-C_3H_6O]^+$  (20), 159  $[M-AcOH-C_3H_6O]^+$  (20), 159  $[M-AcOH-C_3H_6O]^+$  (40), 126 (50), 107 (60), 93 (100), 67 (50), 55 (50), 43 (80).

Ketone 9. Compound 7 (70 mg) was oxidized overnight in 5 ml CH<sub>2</sub>Cl<sub>2</sub> with PDC (70 mg), yielding a product which was chromatographed on silica gel (10 g; hexane-Et<sub>2</sub>O, 8:2) to afford crystalline 9 (30 mg). Mp 127-129° (hexane),  $[\alpha]_D$  -38.9° (CHCl<sub>3</sub>; c 2.3). IR  $v_{max}^{nujol}$  cm<sup>-1</sup>: 2950, 1705, 1460, 1380, 1290, 1230, 1170, 1110, 1070, 1010, 980, 800, 700. EIMS (probe) 70 eV m/z (rel. int.): 234  $[M - C_3H_6O]^*$  (4), 216  $[M - C_3H_4O - H_2O]^*$  (2), 201  $[M - C_3H_4O - Me]^*$  (2), 150 (4), 135 (10), 122 (10), 106 (10), 93 (25), 83 (100), 67 (10), 55 (20).

8,12-Isopropylidene derivative 10. Mp 103-105° (hexane).  $[\alpha]_D$  = 87.5° (CHCl<sub>3</sub>; c 1.08). IR  $v_{max}^{nujol}$  cm<sup>-1</sup>: 3400, 1670, 1390, 1280, 1260, 1220, 1100, 1080, 1020, 960, 890, 850, 830. EIMS (probe) 70 eV m/z (rel. int.): 236  $[M-C_3H_4O]^*$  (5), 218  $[M-C_3H_4O-H_2O]^*$  (10), 159 (15), 122 (25), 107 (35), 93 (100), 84 (80), 69 (40), 55 (40).

Dehydration of 2 to yield furane 13. Product 2 (312 mg) was reacted at room temp, with TsCl (200 mg) in pyridine (6 ml). After

5 hr and usual work up the oily residue was chromatographed on silica gel (20 g) with hexane–Et<sub>2</sub>O (8:2) yielding 13 (46 mg). Oily,  $[\alpha]_D = 36.6^{\circ}$  (CHCl<sub>3</sub>; c 3.5). IR  $v_{\rm max}^{\rm dim}$  cm<sup>-1</sup>: 1720, 1650, 1470, 1400, 1240, 1150, 1040, 980, 850. EIMS (probe) 70 eV m/z (rel. int.): 318 [M]\* (1), 218 [M-AngOH]\* (5), 203 [M-AngOH-Me]\* (3), 110 (30), 93 (30), 91 (30), 85 (60), 83 (100), 55 (60), 43 (20).

12-Angeloxy-8-O-angeloylshiromodiol (14). Oily,  $[\alpha]_D = 38.0^{\circ}$  (CHCl<sub>3</sub>; c 0.9). 1R  $v_{\rm max}^{\rm lim}$  cm<sup>-1</sup>: 3500, 1720, 1660, 1460, 1400, 1360, 1250, 1170, 1100, 1050, 1000, 840. EIMS (probe) 70 eV m/z (rel. int.); 234 (1), 203 (7), 170 (60), 150 (40), 141 (70), 77 (100), 58 (80), 51 (30), 43 (70), 41 (50), 39 (30), 29 (10).

To a soln of 1 (35 mg) in CHCl<sub>3</sub> (1 ml) with a small amount of  $K_2CO_3$ , portions of m-CPB acid were added and the reaction followed by TLC. After usual work up, the crude product was purified by chromatography (silica gel, 10 g, hexane-Et<sub>2</sub>O, 7:3) to give a monoepoxide (25 mg) identical in all respects to 14.

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#### REFERENCES

- Pascual Teresa, J., Pascual, M., Arias, A., Hernández, J. M., Morán, J. R. and Grande, M. (1985) Phytochemistry 24, 1773.
- Pascual Teresa, J., Morán, J. R., Hernández, J. M. and Grande, M. (1985) Phytochemistry 24, 1779.
- Pascual Teresa, J, Morán, J. R., Hernández, J. M. and Grande, M. (1985) Phytochemistry 24, 2071.
- Pascual Teresa, J., Morán, J. R. and Grande, M. (1985) Chem. Letters 865.
- Samek, Z. and Harmatha, J. (1978) Collect. Czech. Chem. Commun. 43, 2779.
- Mc Clure, R. J., Sim, G. A., Goggon, P. and McPhail, A. T. (1970) J. Chem. Soc. Chem. Commun. 128.
- Appendino, G. and Gariboldi, P. (1983) J. Chem. Soc. Perkin Trans. 1, 2017.